

# Recent Advances in the Development of Dopamine D<sub>3</sub> Receptor Agonists and Antagonists

A. Michael Crider<sup>†\*</sup> and Mark A. Scheideler<sup>‡</sup>

<sup>†</sup>Department of Basic Pharmaceutical Sciences, College of Pharmacy, The University of Louisiana at Monroe, Monroe, LA 71209-0470

<sup>‡</sup>Neurobiology Research, Department of Neuroscience, Smithkline Beecham Pharmaceuticals, Via Zambelletti, 20021 Baranzate di Bollate, Milano, Italy

**Abstract:** Advances in molecular cloning techniques have allowed the characterization of five subtypes (D<sub>1</sub>-D<sub>5</sub>) of dopamine (DA) receptors. The limbic location of the D<sub>3</sub> receptor has led to speculation about its possible role in schizophrenia and drug abuse. Since the D<sub>3</sub> receptor is localized in the limbic region rather than the striatum, antipsychotics with D<sub>3</sub> receptor selectivity could be devoid of extrapyramidal side effects commonly seen with D<sub>2</sub> receptor antagonists. Recent work in our laboratory revealed that the benz[e]indole cis-(±)-**44b** demonstrated high selectivity for the D<sub>3</sub> receptor. This compound exhibits a typical antipsychotic profile without the motor effects found in commonly used antipsychotic agents. This mini-review will give a brief introduction on D<sub>3</sub> receptors and a detailed description of selectively-acting D<sub>3</sub> agonists and antagonists which have recently appeared in the literature.

## INTRODUCTION

Advances in molecular techniques have facilitated the design and development of novel ligands with selectivity at dopamine (DA) receptor subtypes. Compounds with selectivity for DA receptors in the limbic region of the brain have been suggested as novel agents in the treatment of schizophrenia and drug abuse [1]. The limbic regions such as the nucleus accumbens, olfactory tubercles, and the islets of Calleja are believed to be involved in thought and emotional processes, whereas the caudate may effect motor control [2]. Thus, compounds that have a selective limbic site of action may lack extrapyramidal side effects of currently available antipsychotic agents [3].

Presently, at least five DA receptor subtypes are known [4]. On the basis of similarities in functional and pharmacological properties, ligand binding, and sequence homologies, these receptors can be divided into two major classes: the D<sub>1</sub>-family (D<sub>1</sub> and D<sub>5</sub>) and the D<sub>2</sub>-family (D<sub>2</sub>, D<sub>3</sub>, and D<sub>4</sub>) [3-5]. Analyses of the amino acid sequences of the DA receptor subtypes reveal that a great deal of receptor homology exists, particularly between members of the same family of DA receptors. The rat and human D<sub>3</sub> receptors contain 400 and 446 amino acids, and the D<sub>3</sub> receptor has overall homologies of 52 % and 41 % with the D<sub>2</sub> and D<sub>4</sub> receptors, respectively. The DA receptor subtypes have been shown to contain seven regions of hydrophobic amino acids fully capable of spanning the cell membrane. Thus, the DA receptors appear to fit the general model of a G-protein coupled receptor with seven membrane spanning  $\alpha$ -helical

regions with extracellular and intracellular loops. The D<sub>1</sub>-family of receptors do not contain introns, whereas the D<sub>2</sub>-family of receptors are interrupted by introns in the gene sequences. The D<sub>2</sub>-family also differs from the D<sub>1</sub>-family with the presence of a long third intracellular loop and a short carboxyl terminal end. The third intracellular loop is believed to be important in the interaction of the G-protein and the receptor. Variants in the D<sub>2</sub> receptor subtype are due to insertion of the 29 amino acids into the third loop to give D<sub>2S</sub> and D<sub>2L</sub> receptor variants [1,5,6]. Variable length polymorphic forms of the D<sub>4</sub> receptor are also known, formed by insertion of repeats of a 48 base pair sequence into the predicted third intracellular loop of the receptor [7].

The D<sub>1</sub> and D<sub>2</sub> receptor subtypes were initially distinguished on the basis of coupling to adenylate cyclase leading to increased and decreased cAMP synthesis, respectively [4]. While D<sub>3</sub> receptors were not initially observed to regulate adenylate cyclase activity, a variety of other cellular responses mediated by this receptor have been observed in heterologous expression systems. These responses include a D<sub>3</sub> receptor-mediated increase in mitogenesis in NG108-15 cells [8] and Na<sup>+</sup>-dependent extracellular acidification in CHO cells [4]. DA agonist-mediated changes in *c-fos* expression have also been seen in NG108-15 cells [9]. Robinson and Caron [10] subsequently showed that the D<sub>3</sub> receptor is able to negatively regulate the type V variant of adenylate cyclase. Several studies have shown that the transmembrane segments (TM) play a critical role in the formation of the ligand binding pocket of catecholamine receptors. The aspartate residue (D110) on TMIII of D<sub>3</sub> receptors probably forms an ionic attraction with the protonated amine group of DA. Using site-directed mutagenesis, the cysteine residue (C114) in TMIII of D<sub>3</sub> receptors was shown to be important in receptor binding of

\*Address correspondence to this author at the Department of Basic

ligands containing *N*-propyl substitution. This cysteine residue is believed to play an important role in maintaining the key "propyl cleft" of the receptor [11].

Although the use of gene knockout and antisense strategies have been utilized, the development of selectively-acting ligands at D<sub>3</sub> receptors is critically important to study the significance and function of this receptor [12]. Depending on the assay conditions, transfected cell line or tissue employed, and radioligand used, the reported affinities and selectivities have varied widely [6]. The binding affinities of a number of DA receptor ligands were recently evaluated at hD<sub>3</sub>, hD<sub>2</sub>, and rat D<sub>2</sub> receptors using several different radioligands [12]. In this study, (+)-7-OH-DPAT exhibited D<sub>2</sub>/D<sub>3</sub> selectivities ranging from 47-302-fold, depending upon the radioligand utilized.

Several functional assays have been utilized to determine agonist or antagonist activity at D<sub>3</sub> receptors. Agonists at D<sub>3</sub> receptors have been shown to enhance mitogenesis in NG108-15 cells [8], increase extracellular acidification in CHO cells [4], and mediate changes in *c-fos* expression in NG108-15 cells [9]. Antagonists at D<sub>3</sub> receptors have been shown to block these effects. However, a number of agonists exhibit higher D<sub>2</sub>/D<sub>3</sub> selectivities in binding studies than in functional tests [4].

Several investigators [6,13,14] have proposed the existence of pre- and postsynaptic D<sub>3</sub> receptors, due to anatomical evidence of postsynaptic receptor expression in the nucleus accumbens and studies suggesting the presynaptic expression of functional receptors in terminals projecting from the substantia nigra. A number of methods have been utilized to assess pre- and postsynaptic activity including measurement of locomotor activity in normal and reserpinized mice, evaluation of contralateral turning in 6-hydroxydopamine (6-OH-DA)-lesioned rats, and determination of intrastitital dopamine levels in rats [15]. The role of DA D<sub>3</sub> receptors in spontaneous locomotor activity has been extensively studied. The work of several groups to profile the activity of selective dopaminergic agents has suggested that agonist action at D<sub>3</sub> receptors inhibits locomotor activity. This is in contrast to the opinion that stimulation of D<sub>2</sub> receptors increases locomotor activity [6,16]. The DA agonists 7-OH-DPAT and PD 128907 were shown to produce a biphasic effect on rat locomotor activity. The inhibition of locomotor activity at lower doses was attributed to D<sub>3</sub> receptor stimulation. At higher doses, however, 7-OH-DPAT leads to an increase in rat locomotor activity, apparently due to D<sub>2</sub> receptor stimulation [16]. In contrast, the D<sub>3</sub> receptor antagonist U99194A was shown to increase spontaneous locomotor activity in mice [13,15]. DA agonists produce contralateral turning in unilateral 6-hydroxydopamine-lesioned rats. This effect is mediated by stimulation of postsynaptic DA receptors. DA D<sub>2</sub> antagonists such as haloperidol block the contralateral turning behavior in rats induced by the D<sub>3</sub> agonist 7-OH-DPAT [3,17]. Maj *et al.* [18] showed that 7-OH-DPAT antagonized catalepsy induced by reserpine, haloperidol, and fluphenazine. The D<sub>3</sub> antagonist nafadotride was shown to antagonize the effect of 7-OH-DPAT. These workers concluded that the anticataleptic action of 7-OH-

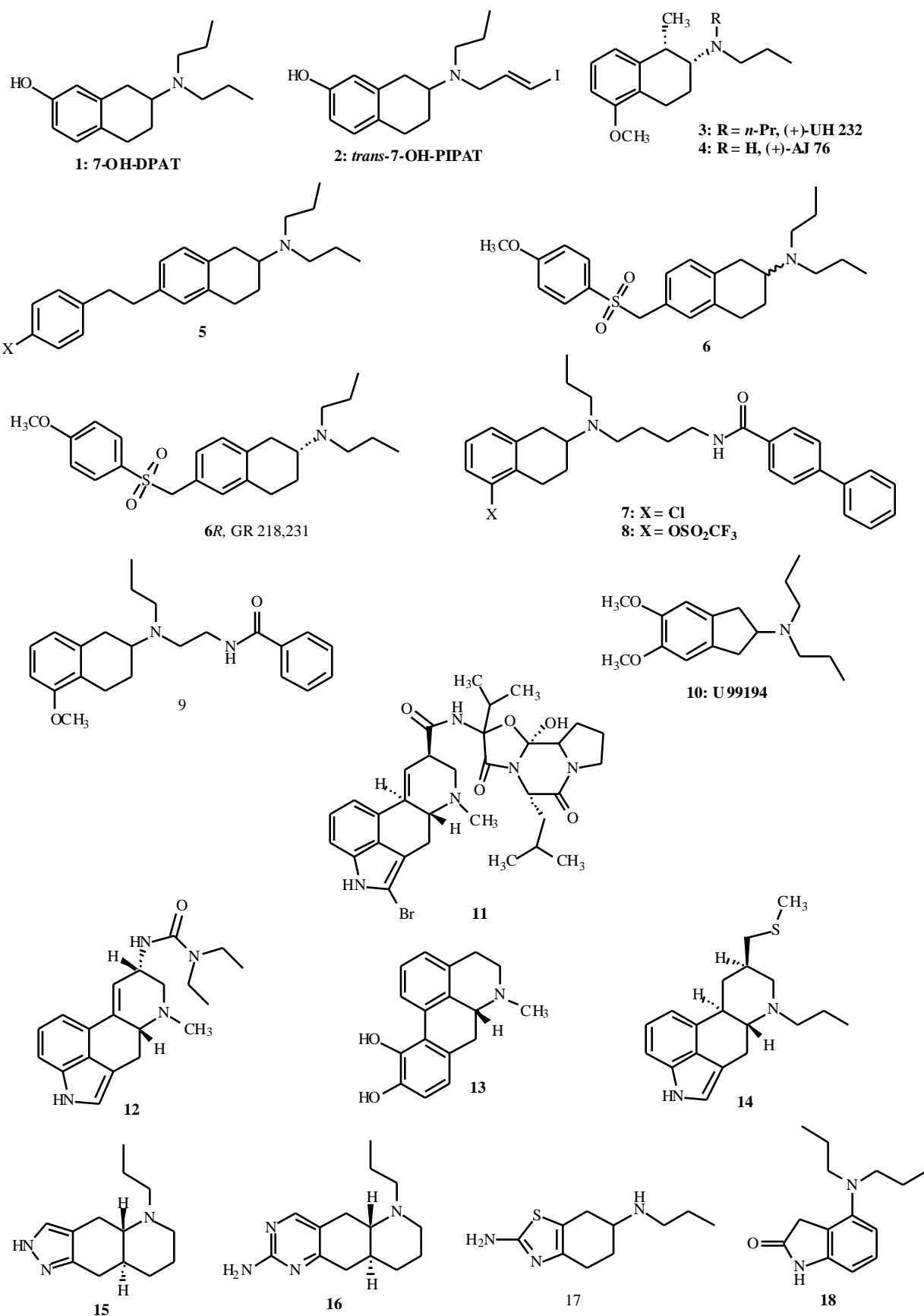
DPAT was due to its action at D<sub>3</sub> receptors. In contrast, Boulay *et al.* [19] used D<sub>3</sub> receptor knock-out mice to show that the D<sub>2</sub> receptor is necessary for haloperidol-induced catalepsy. These workers postulated that D<sub>1</sub>-like, D<sub>2</sub>-like, and D<sub>1</sub>/D<sub>2</sub> antagonists do not involve the D<sub>3</sub> receptor in the production of catalepsy. These studies with receptor-selective ligands and receptor knockout animals suggesting a selective D<sub>3</sub> receptor involvement in regulating cognition and awareness states, and not generalized motor coordination, have suggested a possible therapeutic utility of D<sub>3</sub> agonists in treating Attention Deficit Disorders. However, an initial clinical study examining association with polymorphisms in the D<sub>3</sub> receptor gene has failed to establish a genetic linkage to this disease [20].

## DA D<sub>3</sub> RECEPTOR LIGANDS

The aminotetralin 7-hydroxy-2-(*N,N*-di-*n*-propylamino) tetralin (**1**, 7-OH-DPAT) was the first compound to show preference for the DA D<sub>3</sub> receptor [21,22]. Most of the D<sub>3</sub> receptor binding affinity is found in the *R*(+)-enantiomer [21]. The apparent selectivity of *R*(+)-7-OH-DPAT in radioligand binding studies is greatly decreased when in vivo functional tests are employed to evaluate D<sub>3</sub> receptor activation [23,24]. In fact, several in vivo effects of 7-OH-DPAT are believed to be mediated by D<sub>2</sub> receptors [1]. Receptor binding was initially studied in CHO-K1 cells using [<sup>3</sup>H]spiperone as the radioligand; however, these studies were carried out using conditions which favor binding to GTP-insensitive D<sub>2L</sub> receptor sites with low affinity for agonists. When these studies were performed using the agonist ligand [<sup>3</sup>H]N-0437, only binding to the high affinity site of the D<sub>2L</sub> receptor was measured. Under these conditions, the D<sub>3</sub> versus D<sub>2</sub> selectivity of 7-OH-DPAT was only 24-fold [2]. A recent report showed that 7-OH-DPAT-induced hypothermia and penile erections (PE) were inhibited by D<sub>3</sub>-selective antagonists [25]. Another recent study [26] demonstrated that 7-OH-DPAT decreased intraocular pressure (IOP) in sympathetically-denervated rabbits. This effect was inhibited by D<sub>3</sub> receptor antagonists. These workers suggest that the IOP-lowering effect of 7-OH-DPAT is due to stimulation of postsynaptic D<sub>3</sub> receptors located on sympathetic nerve endings in the ciliary body.

An iodinated analog of 7-OH-DPAT, (*R,S*-2'-*trans*-7-hydroxy-2-[*N-n*-propyl-*N*-(3'-iodo-2'-propenyl)amino]tetralin (*trans*-7-OH-PIPAT, **2**) was reported to exhibit a 143-fold D<sub>3</sub> versus D<sub>2</sub> selectivity [27]. Binding studies using membrane preparations containing D<sub>3</sub> receptors expressed in *Spodoptera frugiperda* (Sf9) cells showed that the activity resides with the *R*-enantiomer [28]. Further studies demonstrated that 7-OH-PIPAT additionally binds with high affinity at 5HT<sub>1A</sub> [28] and -receptors [29].

The 5-methoxy-2-aminotetralin (**3**, (+)-UH 232) was shown to bind with a 4-fold D<sub>3</sub> versus D<sub>2</sub> selectivity [30]. The pharmacological profile of compound **3** and the related *n*-propyl derivative (**4**, (+)-AJ 76) suggests that both of these compounds act as DA autoreceptor antagonists [31,32]. However, more recent reports indicate that UH 232 may be acting as a partial agonist [8].

**Fig. (1).** Structures of D<sub>3</sub> ligands (1-18).

Using (+)-UH 232 as a structural lead, Murray *et al.* [33] prepared a series of 6-hydroxy-2-aminotetralins with high binding affinity for D<sub>3</sub> receptors. A number of compounds of general structure **5**, in which the 5-methoxy group of (+)-UH 232 was removed, exhibited high binding affinity for the D<sub>3</sub> receptor with greater than 100-fold D<sub>3</sub> versus D<sub>2</sub> selectivity. The high lipophilicity of the aminotetralins **5** make clinical development of these compounds unlikely due to rapid *in vivo* clearance. As a result, several derivatives with decreased lipophilicity were prepared. The sulfone **6** exhibited high binding affinity and favorable D<sub>3</sub>/D<sub>2</sub> selectivity. The *R*-enantiomer (*R*-**6**, GR 218,231) was shown to be the active isomer, and the compound demonstrated D<sub>3</sub>/D<sub>2</sub> selectivity of 400-fold and about 10,000-fold D<sub>3</sub> versus D<sub>1</sub> or D<sub>4</sub> selectivity. A report by Cussac *et al.* [34] showed that GR 218,231 displaced [<sup>3</sup>H]7-OH-DPAT with similar affinities at cloned and native rat D<sub>3</sub> receptors. The affinity of GR 218,231 at these sites was much higher than at D<sub>2</sub> receptors in the striatum. GR 218,231 dose-dependently reduced 7-OH-DPAT-induced PE and hypothermia, but was inactive against 7-OH-DPAT-induced yawning and hypothermia. Furthermore, GR 218,231 did not block contralateral turning behavior caused by quinpirole. These workers also demonstrated that GR 218,231 was inactive in models to predict antipsychotic and extrapyramidal activity. This study further confirms the need for development of selectively-acting ligands to determine the role of D<sub>2</sub> and D<sub>3</sub> receptors in the study and treatment of schizophrenia.

A series of 5-substituted-*N*-*n*-propyl-2-aminotetralins were recently described by Boyfield *et al.* [35]. Compounds **7** and **8** demonstrated high binding affinity for the DA D<sub>3</sub> receptor (K<sub>i</sub> = 2.2 nM) with about 200-fold D<sub>3</sub> versus D<sub>2</sub> selectivity. These compounds were shown, however, to undergo rapid clearance due to *N*-depropylation.

Homan *et al.* [36] prepared a series of compounds which were structural hybrids of the aminotetralins and the 2-pyrrolidinylmethylbenzamides. The lead compound (5-methoxy-2-[*N*-(2-benzamidoethyl)-*N*-(*n*-propyl)amino]tetralin, **9**) exhibited binding affinities at 5HT<sub>1A</sub>, D<sub>2A</sub>, and D<sub>3</sub> receptors of 0.82 nM, 3.2 nM, and 0.58 nM, respectively. These workers postulated that the benzamidoethyl side chain binds to an accessory site at all three receptors, and this site may be identical to that occupied by the 2-pyrrolidinylmethylbenzamide D<sub>2</sub>/D<sub>3</sub> antagonists. Furthermore, these workers speculated that these antagonists at 5-HT<sub>1A</sub> and D<sub>2</sub>/D<sub>3</sub> receptors may have a reduced tendency to produce extrapyramidal side effects (EPS).

The preferential D<sub>3</sub> antagonist U 99194 (**10**, U 99194) was shown to increase rat locomotor activity through postsynaptic D<sub>3</sub> receptor stimulation [37]. Using various radioligands to label the D<sub>3</sub> receptor, Audinot *et al.* [12] reported K<sub>i</sub> values for U 99194 of between 160-223 nM with D<sub>3</sub>/D<sub>2</sub> selectivity of 10 to 14-fold. Additionally, these workers showed that U 99194 antagonized the induction of hypothermia by (+)-7-OH-DPAT and did not induce catalepsy in rats.

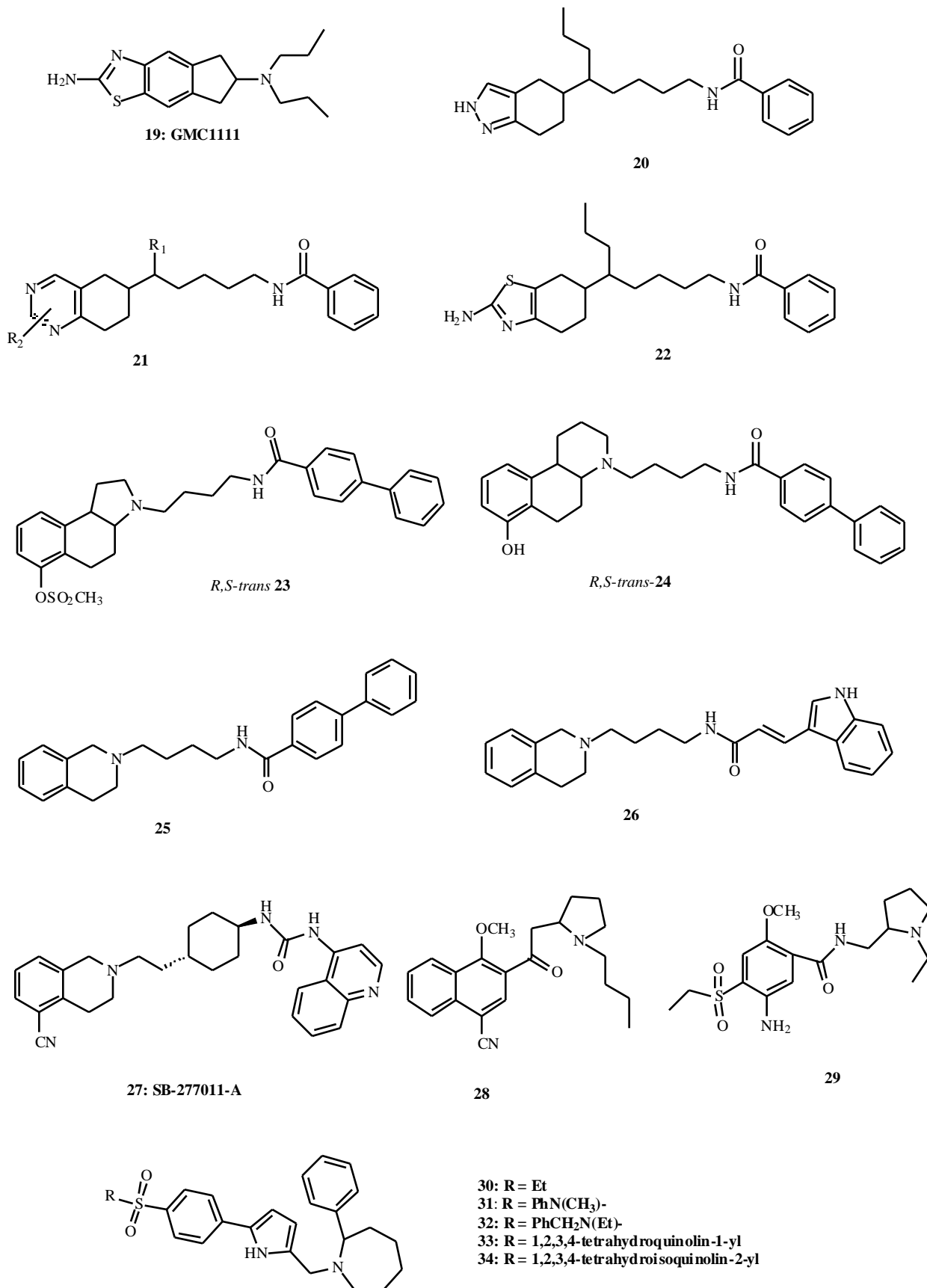
In displacement studies and functional tests based on the mitogenic response to D<sub>3</sub> or D<sub>2</sub> receptor stimulation, Griffon *et al.* [30] showed that the ergolines, bromocriptine (**11**) and

lisuride (**12**), and apomorphine (**13**) exhibited high binding affinities at both subtypes with no apparent D<sub>3</sub>/D<sub>2</sub> selectivity. Pergolide (**14**), an ergoline with antiparkinsonian activity, exhibited high binding affinities at both D<sub>2</sub> and D<sub>3</sub> receptors with about an 8-fold selectivity at D<sub>3</sub> receptors. Pergolide was also shown to be a full agonist in the mitogenic assay [38].

The tricyclic analogs quinpirole (**15**) and quinolorane (**16**) show selectivities at D<sub>3</sub> versus D<sub>2</sub> receptors in radioligand binding experiments of 36- and 95-fold, respectively [4]. However, in the mitogenic assay to measure functional activity only quinolorane exhibited some selectivity (approximately 20-fold). The related bicyclic aminothiazole derivative pramipexole (**17**) binds with high affinity at both D<sub>2</sub> and D<sub>3</sub> receptors with about a 70-fold selectivity for D<sub>3</sub> receptors. In the mitogenic response in cells expressing D<sub>2</sub> or D<sub>3</sub> receptors, pramipexole demonstrated over 80-fold D<sub>3</sub> selectivity [38]. Pramipexole inhibits DA synthesis and release and neuronal firing by acting on DA autoreceptors [39]. The 2-indolone ropinirole (**18**) demonstrated a 16-fold D<sub>3</sub>/D<sub>2</sub> selectivity with full agonist activity in the mitogenic functional assay [38]. Both pramipexole and ropinirole have been used as monotherapy and in combination with levodopa in the treatment of parkinsonism [40,41]. The efficacy of these compounds may be related to their high D<sub>3</sub> binding affinity [38]. However, several studies have suggested action via activation of D<sub>2</sub> autoreceptors as a likely mechanism of drug action [42,43]. This would be consistent with the high D<sub>2</sub> receptor expression and relative paucity of D<sub>3</sub> receptors in motoric regions of the brain, and D<sub>2</sub> agonist activity of these compounds [44].

Incorporation of the 2-aminothiazole moiety into the 2-aminoindane nucleus led to the discovery that (**19**, GMC 1111) had high affinity for D<sub>3</sub> receptors. This compound caused rotation in 6-hydroxydopamine (6-OH-DA)-lesioned rats and an increase in DA turnover in the striatum. In the mitogenic assay to assess functional activity, GMC 1111 was shown to be a partial agonist at D<sub>2</sub> receptors and an antagonist at D<sub>3</sub> receptors. Additionally, GMC 1111 demonstrated good oral bioavailability in rats [45]. These workers suggested that GMC 1111 may have potential therapeutic utility in the treatment of Parkinson's disease, as suggested by the combined D<sub>3</sub> antagonist and D<sub>2</sub> agonist activity of the compound.

Incorporation of the 4-(4-phenylbenzoylamino)butyl side-chain, found in compounds **7** and **8**, into structural fragments of quinpirole, quinolorane, and pramipexole led to the synthesis of the corresponding bicyclic derivatives (**20-22**) [46]. These compounds were evaluated in displacement studies using [<sup>125</sup>I]iodosulpride for D<sub>2</sub> and D<sub>3</sub> receptors expressed in CHO cells. Functional activity was determined by microphysiometry. The quinolorane analog (**21**, R<sub>2</sub> = 4-OCH<sub>3</sub>, R<sub>1</sub> = *n*-Pr) showed high D<sub>3</sub> receptor affinity with 490-fold D<sub>3</sub>/D<sub>2</sub> selectivity. The quinpirole (**20**) and pramipexole derivative (**22**) analogs also demonstrated high D<sub>3</sub> binding affinities and D<sub>3</sub> versus D<sub>2</sub> selectivities of 150-fold and 340-fold, respectively. These derivatives were shown to act as agonists in the functional assay.

Fig. (2). Structures of D<sub>3</sub> ligands (19-34).

The aminotetralins, as typified by compounds **7** and **8**, were shown to be metabolized by *N*-dealkylation of the *n*-propyl group and undergo rapid clearance. In an attempt to improve the metabolic stability of these analogs, the *N*-propyl group was incorporated into either a benz[*e*]indole or benzof[*l*]quinoline nucleus. The ( $\pm$ )-*trans*-benz[*e*]indole **23** and the ( $\pm$ )-benzof[*l*]quinoline **24** were shown to bind with high affinity at D<sub>3</sub> receptors with D<sub>3</sub>/D<sub>2</sub> selectivities of 65-fold and 72-fold, respectively. Using microphysiometry, compounds **23** and **24** acted as antagonists at D<sub>3</sub> receptors. These tricyclic analogs had improved *in vivo* stability compared to the aminotetralins [47]. These same workers used brain penetration and clearance studies to develop a novel isoquinoline **25** with high D<sub>3</sub> affinity and good *in vivo* stability. Modification of compound **25** led to the *N*-(3-indolylpropenamido) derivative **26**, a compound with high affinity for the D<sub>3</sub> receptor ( $K_i = 4$  nM) and 150-fold D<sub>3</sub>/D<sub>2</sub> selectivity. Similar to the isoquinoline **25**, compound **26** demonstrated a low blood clearance and CNS penetration following IV administration. The plasma half-life of compound **26** was 2.5 hours [48]. Further studies on compound **26** demonstrated oral bioavailability of only 7 % and modest affinity for the 5HT<sub>1B</sub> and 5HT<sub>1D</sub> receptors. Replacement of the 7-trifluorosulfonyl group with a cyano-substituent slightly improved oral bioavailability, but reduced D<sub>3</sub> receptor affinity. Molecular modeling studies suggested that the flexible butyl spacer in compound **26** may impart 5HT<sub>1B</sub> and 5HT<sub>1D</sub> receptor binding affinity. Using this information, a number of conformationally constrained linkers were investigated. Further structural modification yielded the 4-quinolinyl derivative (**27**, SB-277011). This compound exhibited high D<sub>3</sub> receptor affinity with over 100-fold selectivity against D<sub>2</sub>, 5HT<sub>1B</sub>, and 5HT<sub>1D</sub> receptors. Additionally, this analog demonstrated D<sub>3</sub> antagonist activity, good oral bioavailability (43 %), low blood clearance, and high CNS activity. The compound does not induce catalepsy or elevate prolactin levels. Additional studies using microphysiometry revealed that SB-277011-A antagonized the effect of quinpirole-induced extracellular acidification rates with over 80-fold D<sub>3</sub>/D<sub>2</sub> selectivity in CHO cells. Additionally, SB-277011-A did not alter spontaneous locomotion or enhance amphetamine or phencyclidine-induced hyperactivity *per se*, and significantly reversed a prepulse inhibition deficit in isolation-reared rats [49]. Thus, this novel isoquinoline may have a reduced tendency to produce extrapyramidal side effects which are typical of classical antipsychotics such as haloperidol [50].

Sautel *et al.* [51] recently described the pharmacological profile of nafadotride (**28**) and its enantiomers. In displacement studies, the (-)-isomer displays about 20-fold greater affinity for the D<sub>3</sub> receptor ( $K_i = 0.3$  nM) than the (+)-isomer. The (+)-isomer shows only 2-fold D<sub>3</sub>/D<sub>2</sub> selectivity; however, the (-)-isomer exhibits about 10-fold D<sub>3</sub> selectivity. Nafadotride failed to show any efficacy in the mitogenic response of the DA agonist quinpirole. At low doses (0.1-1.0 mg/kg) nafadotride, unlike haloperidol, increased rat spontaneous locomotor activity and climbing behavior in mice. Similar to haloperidol, nafadotride induced catalepsy at high doses (1-100 mg/kg). These researchers suggested that the increase in motor activity was due to D<sub>3</sub> receptor blockade. Griffon *et al.* [30] postulated that activation of a subpopulation of mesolimbic D<sub>3</sub> receptors inhibits rat

locomotor activity. Thus, the increase in locomotor activity elicited by nafadotride is consistent with preferential D<sub>3</sub> receptor antagonism. Similar to haloperidol, nafadotride induced catalepsy at high doses (1-100 mg/kg). These researchers suggested that the increase in motor activity was due to D<sub>3</sub> receptor blockade, although this result may reflect D<sub>2</sub> receptor antagonism at the doses employed in this study.

Amisulpride (**29**) is a benzamide derivative that exhibits high binding affinity at D<sub>2</sub> ( $K_i = 2.8$  nM) and D<sub>3</sub> ( $K_i = 3.2$  nM) receptors. In the rat, amisulpride exhibits an antagonist action at pre- and postsynaptic D<sub>2</sub> receptors. At low doses, the compound selectively blocks DA autoreceptors that regulate synthesis and release. This unique pharmacological profile has been suggested to be responsible for the use of amisulpride for the negative symptoms of schizophrenia at low doses and the positive symptoms at high doses. Due to the low incidence of EPS, the clinical use of amisulpride is of considerable interest [52].

The 2,5-disubstituted-1*H*-pyrrole (**30**) was shown to bind with high affinity at D<sub>3</sub> receptors with about a 30-fold selectivity over D<sub>2</sub> receptors [53]. Modification of the ethylsulfone group of **30** yielded several derivatives (**31-34**) with high affinities and D<sub>3</sub> versus D<sub>2</sub> selectivities of over 100-fold [54]. Little enantiomeric selectivity was noted with the isomers of compound **31**, although the *R*-isomer showed higher binding affinities at both D<sub>2</sub> and D<sub>3</sub> receptors.

The tricyclic 1,4-oxazine (**35**, (+)-PD 128,907) was reported by Patel *et al.* [55] to exhibit high binding affinity ( $K_i = 1.7$  nM) and over 100-fold D<sub>3</sub>/D<sub>2</sub> selectivity. Using a mitogenic assay to assess functional activity, (+)-PD 128,907 was found to act as an agonist with approximately 50-fold selectivity at D<sub>3</sub> receptors [4]. Studies by Whetzel *et al.* [56] demonstrated that (+)-PD 128,907 decreased striatal and mesolimbic L-dihydroxyphenylalanine (L-DOPA) levels in rats pretreated with the L-aromatic amino acid decarboxylase inhibitor NSD 1015. This study adds further evidence for the regulatory role of D<sub>3</sub> receptors on the synthesis and release of DA. Millan *et al.* [57] demonstrated that the decrease in rectal temperature in rats induced by 7-OH-DPAT could be modulated by (+)-PD 128,907. These workers suggested that the modulation of hyperthermia in rats involves D<sub>3</sub> receptor activation. Witkin *et al.* [58] used stereotypic behaviors in mice induced by either the DA agonist, apomorphine, or the NMDA antagonist, dizocilpine, to compare the differences among haloperidol, (+)-PD 128,907, and clozapine. Similar to clozapine, (+)-PD 128,907 blocked stereotyped behavior produced by dizocilpine at about a 12-fold lower dose than stereotypy induced by apomorphine. Also, (+)-PD 128,907 failed to produce catalepsy. The authors suggested that the ability of (+)-PD 128,907 to inhibit dizocilpine-induced stereotypic behaviors may be related to its effect on neuronal pathways involving both DA and glutamate. Recent studies have shown that rats can be trained to discriminate (+)-PD 128,907 from saline [59]. However, the use of subtype selective antagonists suggests the involvement of presynaptic D<sub>2</sub> receptors in this response. Thus, these results indicate that the *in vivo* selectivity of (+)-PD 128,907 is much less than its *in vitro* selectivity. The thiopyran isostere (**36**) of (+)-PD 128,907 was recently reported by van Vliet *et al.*

[60]. The highest binding affinity in the *trans*-series resided with the (-)-isomer. Unlike (+)-PD 128,907, *trans*-**36** did not exhibit any DA receptor selectivity, but demonstrated agonist activity at 5HT<sub>1A</sub> receptors. The *cis*-isomer acted as a full agonist at D<sub>2</sub> receptors and a partial agonist at D<sub>3</sub> receptors with about a 20-fold D<sub>3</sub>/D<sub>2</sub> selectivity.

The benzofuran (±)-S 11566 exhibited preferential affinity for D<sub>3</sub> receptors in displacement studies using [<sup>125</sup>I]iodosulpride as the radioligand. The active isomer was shown to be the (+)-isomer (**37**, S 14297) with D<sub>3</sub>/D<sub>2</sub> K<sub>i</sub> values of 13 nM and 297 nM and over 20-fold D<sub>3</sub>/D<sub>2</sub> selectivity [61]. Additionally, (+)-S 14297 inhibited hypothermia induced by a variety of D<sub>3</sub> agonists including 7-OH-DPAT and (-)-quinpirole. In this study, potency for

inhibition of 7-OH-DPAT-induced hypothermia correlated well with D<sub>3</sub> receptor binding affinity. Unlike nonselective DA antagonists such as haloperidol, (+)-S 14297 did not produce catalepsy or stimulate prolactin release. However, (+)-S 14297 demonstrated partial agonist activity in stimulation of D<sub>3</sub>-coupled mitogen-activated protein kinase (MAPK) [62].

The chromanopyrrole (**38**, S 33084) was shown to act as preferential D<sub>3</sub> antagonist with greater than 100-fold D<sub>3</sub>/D<sub>2</sub> selectivity and greater than 100-fold selectivities at over 30 other receptors evaluated [63]. Although S 33084 inhibited hypothermia and PE induced by 7-OH-DPAT and (+)-PD 128,907, the compound was inactive in several models used to predict antipsychotic or extrapyramidal activities.

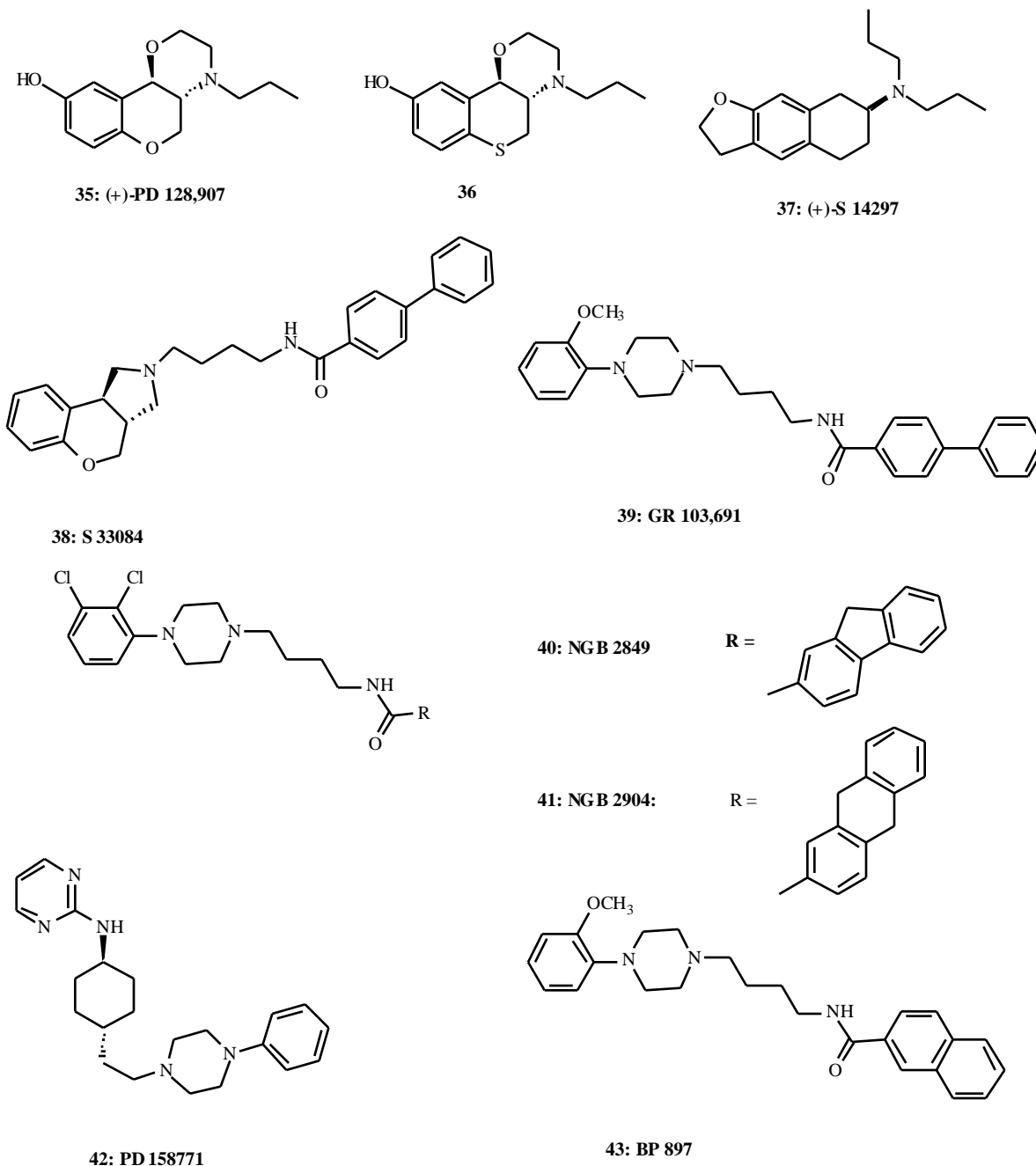
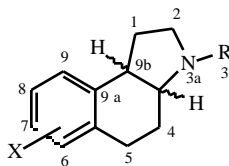


Fig. (3). Structures of D<sub>3</sub> ligands (19-34).

Table 1. In Vitro Binding at D<sub>2S</sub> and D<sub>3</sub> Receptors<sup>a,b</sup>

Compound	R	X	B/C Ring Junction	D <sub>2S</sub> <sup>c</sup>	D <sub>3</sub> <sup>d</sup>	D <sub>2S</sub> /D <sub>3</sub> <sup>e</sup>
(±)-44a	<i>n</i> -Pr	8-OCH <sub>3</sub>	<i>cis</i>	5070±357	1500±158	3
(±)-44b	<i>n</i> -Pr	8-OH	<i>cis</i>	1576±237	39.2±3.0	40
(+)-44b	<i>n</i> -Pr	8-OH	<i>cis</i> <sup>f</sup>	5476±1194	296±12	19
(-)-44b	<i>n</i> -Pr	8-OH	<i>cis</i> <sup>g</sup>	1007±45	14.7±4.3	69
(±)-44c	allyl	8-OCH <sub>3</sub>	<i>cis</i>	3630±201	553±0.5	7
(±)-44d	allyl	8-OH	<i>cis</i>	2907±34	10±1	290
(±)-44e	<i>n</i> -Pr	8-OCH <sub>3</sub>	<i>trans</i>	8417±1338	368±66	23
(±)-44f	<i>n</i> -Pr	8-OH	<i>trans</i>	147±21	2.1±1.9	73
(+)-44f	<i>n</i> -Pr	8-OH	<i>trans</i> <sup>h</sup>	105±21	2.3±1.9	46
(-)-44f	<i>n</i> -Pr	8-OH	<i>trans</i> <sup>i</sup>	2100±569	156±2.5	14
(±)-44g	allyl	8-OCH <sub>3</sub>	<i>trans</i>	2760±429	158±43	18
(±)-44h	allyl	8-OH	<i>trans</i>	57±4	1.7±0.2	34
34				202±123	0.4±0.3	505
14				80±42	1.2±0.8	67

<sup>a</sup>Competitive displacement studies were performed using membrane preparations from clonal cells expressing D<sub>2S</sub> (LtK<sup>-</sup> cells) and D<sub>3</sub> (BHK 21tK<sup>-</sup> cells) receptors, respectively. <sup>b</sup>K<sub>i</sub> values were reported earlier [65,66]. <sup>c</sup>[<sup>3</sup>H]Spiperone (0.1 nM) was used to label D<sub>3</sub> receptors. <sup>d</sup>[<sup>3</sup>H]R-(+)-7-OH-DPAT (0.1 nM) was used to label D<sub>2</sub> receptors. <sup>e</sup>D<sub>3</sub> selectivity, D<sub>2S</sub>(K<sub>i</sub>)/D<sub>3</sub> (K<sub>i</sub>). <sup>f</sup>Absolute configuration (3a*S*,9b*R*). <sup>g</sup>Absolute configuration (3a*R*,9b*S*). <sup>h</sup>Absolute configuration (3a*R*,9b*R*). <sup>i</sup>Absolute configuration (3a*S*,9b*S*).

Furthermore, S 33084 did not enhance locomotor activity or inhibit yawning behavior induced by 7-OH-DPAT. These authors concluded that only the induction of hypothermia and PE involve D<sub>3</sub> receptors [25].

The arylpiperazine (**39**, GR 103,691) was shown to bind with high affinity at the D<sub>3</sub> receptor [64]. In comparison studies with (+)-S 14297, nafadotride, U 99194, and GR 103,691 exhibited a K<sub>i</sub> = 0.40 nM and a D<sub>3</sub>/D<sub>2</sub> selectivity of 60-fold. However, GR 103,691 exhibits significant affinity for 5HT<sub>1A</sub> and <sub>1</sub>-adrenergic receptors. Furthermore, GR 103,691 demonstrates little in vivo bioavailability and appears to only be useful as an in vitro pharmacological tool [12]. Yuan *et al.* [65] showed that the 2,3-dichlorophenylpiperazines (**40**, NGB 2849) and (**41**, NGB 2904) demonstrated high D<sub>3</sub> receptor binding affinities (K<sub>i</sub> = 0.9 nM and 1.4 nM, respectively) and selectivities of over 150-fold versus other DA receptor subtypes. In a mitogenic assay to evaluate functional activity, NGB 2849 and NGB 2904 inhibited quinpirole-stimulated mitogenesis with IC<sub>50</sub> values of 8.8 and 6.8 nM, respectively. The aminopyrimidine (**42**, PD 158771) was found to possess high affinity for D<sub>2</sub>, D<sub>3</sub>, and 5HT<sub>1A</sub> receptors [66]. Further evaluation of this compound revealed partial agonism at DA receptors and agonist activity at 5HT<sub>1A</sub> receptors [67,68].

Pilla *et al.* [69] reported that the D<sub>3</sub> partial agonist (**43**, BP 897) inhibited cocaine-seeking behavior in rats. These workers suggested that BP 897 could be potentially useful for inhibiting cocaine craving and relapse susceptibility produced by drug-related environmental stimuli.

Scheideler *et al.* [70] reported that the benz[e]indole (**44b**, *cis*-(±)-8-OH-PBZI) binds with high affinity for D<sub>3</sub> receptors with selectivities of 775-fold, 550-fold, 90-fold, and 10-fold at D<sub>1A</sub>, D<sub>5</sub>, D<sub>2S</sub>, and D<sub>4</sub> receptors. The compound demonstrated agonist activity at D<sub>3</sub> receptors with a selective limbic site of action. Additional behavioral and neurochemical studies were performed to fully characterize the pharmacological profile of *cis*-(±)-8-OH-PBZI [15]. This compound decreased spontaneous locomotor activity in mice, inhibited conditioned avoidance responding in rats, and failed to elicit catalepsy. At doses which inhibited spontaneous locomotor activity in rats, *cis*-(±)-8-OH-PBZI did not alter interstitial levels of DA or DOPAC in the nucleus accumbens or dorsolateral striatum. The pharmacological profile of *cis*-(±)-8-OH-PBZI suggests an action at postsynaptic D<sub>3</sub> receptors with possible antipsychotic potential devoid of undesirable motor side effects. Using *cis*-(±)-8-OH-PBZI as the structural lead, a detailed structure-activity relationship (SAR) study was



performed (Table 1) [71]. The *trans*-diastereoisomers exhibited greater binding affinities for DA D<sub>3</sub> receptors than the corresponding *cis*-isomers. In both series, the greatest binding affinities were found with compounds substituted with either *n*-propyl or allyl groups at the 3-position of the benz[e]indole nucleus. The allyl derivative *cis*-(±)-**44d** exhibited the greatest D<sub>3</sub>/D<sub>2</sub> selectivity (290-fold) of any benz[e]indole evaluated in this study. Resolution of *cis*-(±)-**44b** or *trans*-(±)-**44f** into individual isomers showed that in both series the more active isomer had 3aR absolute configuration.

## SUMMARY

In about one decade since its discovery, a vast amount of research has occurred on the study of the D<sub>3</sub> receptor and the development of selectively-acting D<sub>3</sub> agonists and antagonists. Potential therapeutic targets for these novel agents include Parkinson's disease, schizophrenia, and cocaine abuse. The continued development of selectively-acting ligands will greatly assist in determining the functional role for this novel receptor and its significance in numerous disease states.

## REFERENCES

- [1] Levant, B. *Pharmacol. Rev.* **1997**, *49*, 231-252.
- [2] Pugsley, T.A.; Davis, M.D.; Akunne, H.C.; MacKenzie, R.G.; Shih, Y.H.; Wikstrom, H.; Whetzel, S.Z.; Georgic, L.M.; Cooke, L.W. *J. Pharmacol. Exp. Ther.* **1995**, *275*, 1355-1366.
- [3] McElroy, J.F.; Ward, K.A.; *Drug Dev. Res.* **1995**, *34*, 329-335.
- [4] Sokoloff, P.; Schwartz, J.-C. *TiPS* **1995**, *16*, 270-274.
- [5] Griffon, N.; Diaz, J.; Levesque, D.; Sautel, F.; Schwartz, J.-C.; Sokoloff, P.; Simon, P.; Costentin, J.; Garrido, F.; Mann, A.; Wermuth, C. *Clin. Pharmacol.* **1995**, *18* (Suppl.), S130-142.
- [6] Shafer, R.A.; Levant, B. *Psychopharmacology* **1998**, *135*, 1-16.
- [7] Van Tol, H.H.M.; Wu, C.M.; Guan, H.C.; Ohara, K.; Bunzow, J.R.; Civelli, O.; Kennedy, J.; Seeman, P.; Niznik, H.B.; Jonanovic, V. *Nature* **1992**, *358*, 149-152.
- [8] Griffon, N.; Pilon, C.; Schwartz, J.-C.; Sokoloff, P. *Eur. J. Pharmacol.* **1995**, *282*, R3-R4.
- [9] Pilon, C.; Levesque, D.; Dimitriadou, V.; Griffon, N.; Martres, M.-P.; Schwartz, J.-C.; Sokoloff, P. *Eur. J. Pharmacol.* **1994**, *268*, 129-139.
- [10] Robinson, S.W.; Caron, M.G. *Mol. Pharmacol.* **1997**, *52*, 508-514.
- [11] Alberts, G.L.; Pregenzer, J.F.; Im, W.B. *Br. J. Pharmacol.* **1998**, *125*, 705-710 and references cited therein.
- [12] Audinot, V.; Newman-Tancredi, A.; Gobert, A.; Rivet, J.-M.; Brocco, M.; LeJune, F.; Gluck, L.; Desposte, I.; Bervoets, K.; DeKeyne, A.; Millan, M.; *J. Pharmacol. Exp. Ther.* **1998**, *287*, 187-197.
- [13] Waters, N.; Svensson, K.; Haadma-Svensson, S.R.; Smidt, M.W.; Carlson, A. *J. Neural Transm.* **1993**, *94*, 11-19.
- [14] Levesque, D.; Martres, M.P.; Diaz, J.; Griffon, N.; Lammers, C.H.; Sokoloff, P.; Schwartz, J.-C. *Proc. Natl. Acad. Sci. USA* **1995**, *92*, 1719-1723.
- [15] Fink-Jensen, A.; Nielsen, E.B.; Hansen, L.; Scheideler, M.A. *Eur. J. Pharmacol.* **1998**, *342*, 153-161.
- [16] Svensson, K.; Carlsson, A.; Waters, N. *J. Neural Transm.* **1994**, *95*, 71-74.
- [17] McElroy, J.F.; Zeller, K.; Amy, K.; Ward, K.; Cawley, J.; Mazzola, A.; Keim, W.; Rohrbach, K.; *Drug Dev. Res.* **1993**, *30*, 257-259.
- [18] Maj, J.; Rogoz, Z.; Skuza, G. *J. Neural Transm.* **1999**, *106*, 1063-1073.
- [19] Boulay, D.; Depoortere, R.; Oblin, A.; Sanger, D.J.; Shoemaker, H.; Perrault, G. *Eur. J. Pharmacol.* **2000**, *391*, 63-73.
- [20] Barr, C.I.; Wigg, K.G.; Wu, J.; Zai, C.; Bloom, S.; Tannock, R.; Roberts, W.; Malone, M.; Schachar, R.; Kennedy, J.L. *Am. J. Med. Genet.* **2000**, *96*, 114-117.
- [21] Levesque, D.; Diaz, J.; Pilon, C.; Martres, M.P.; Giros, B.; Souil, E.; Schott, D.; Morgat, J.L.; Schwartz, J.-C.; Sokoloff, P. *Proc. Natl. Acad. Sci. USA* **1992**, *89*, 8155-8159.
- [22] Damsma, G.; Botlema, T.; Westerink, B.H.C.; Tepper, T.G.; Dijkstra, D.; Pugsley, T.A.; MacKenzie, R.G.; Heffner, T.G.; Wikstrom, H. *Eur. J. Pharmacol.* **1993**, *249*, R9-R10.
- [23] Sautel, F.; Griffon, N.; Levesque, D.; Pilon, C.; Schwartz, J.-C.; Sokoloff, P. *Neuroreport* **1995**, *6*, 329-332.
- [24] Levesque, D. *Biochem. Pharmacol.* **1996**, *52*, 511-518.
- [25] Millan, M.J.; DeKeyne, A.; Rivet, J.-M.; Dubuffet, T.; Lavielle, G.; Brocco, M. *J. Pharmacol. Exp. Ther.* **2000**, *293*, 1063-1073.
- [26] Chu, E.; Chu, T.-C.; Potter, D.E. *J. Pharmacol. Exp. Ther.* **2000**, *293*, 710-716.
- [27] Foulon, C.; Kung, M.-P.; Kung, H.F. *J. Med. Chem.* **1993**, *36*, 1499-1500.
- [28] Burris, K.D.; Filtz, T.; Chumpradit, S.; Kung, M.-P.; Foulon, C.; Hensler, J.G.; Kung, H.F.; Molinoff, P.B. *J. Pharmacol. Exp. Ther.* **1994**, *268*, 935-942.
- [29] Garner, S.E.; Kung, M.-P.; Foulon, C.; Chumpradit, S.; Kung, H.F. *Life Sci.* **1994**, *54*, 593-603.
- [30] Griffon, N.; Sokoloff, P.; Diaz, J.; Levesque, D.; Sautel, F.; Schwartz, J.-C.; Simon, P.; Costentin, J.; Garrido, F.; Mann, A.; Wermuth, C. *Eur. Neuropsychopharmacol. Suppl.* **1995**, 3-9.
- [31] Johansson, A.M.; Arvidsson, L.-E.; Hacksell, U.; Nilsson, J.L.G.; Svensson, K.; Hjorth, S.; Clark, D.; Carlsson, A.; Sanchez, D.; Andersson, B.; Wikstrom, H.; *J. Med. Chem.* **1985**, *28*, 1049-1053.

- [32] Svensson, K.; Johansson, A.M.; Magnusson, T.; Carlsson, A. *Naunyn-Schmiedberg's Arch. Pharmacol.* **1986**, *334*, 234-235.
- [33] Murray, P.J.; Helden, R.M.; Johnson, M.R.; Robertson, G.M.; Scopes, D.I.C.; Stokes, M.; Wadman, S.; Whitehead, J.W.F.; Hayes, A.G.; Kilpatrick, G.J.; Large, C.; Stubbs, C.M.; Turpin, M.P. *Bioorg. Med. Chem. Lett.* **1996**, *6*, 403-408.
- [34] Cussac, D.; Newman-Tancredi, A.; Sezgin, L.; Millan, M.J. *Eur. J. Pharmacol.* **2000**, *394*, 47-50.
- [35] Boyfield, I.; Coldwell, M.C.; Hadley, M.S.; Johnson, C.N.; Riley, G.J.; Scott, E.E.; Stacey, R.; Stemp, G.; Thewlis, K.M. *Bioorg. Med. Chem. Lett.* **1997**, *7*, 1995-1998.
- [36] Homan, E.J.; Copinga, S.; Elfstrom, L.; van der Veen, T.; Hallema, J.-P.; Mohell, N.; Unelius, L.; Johansson, R.; Wikstrom, H.V.; Grol, C.J. *Bioorg. Med. Chem.* **1998**, *6*, 2111-2116.
- [37] Waters, N.; Svensson, K.; Haadsma-Svensson, S.R.; Smith, M.W.; Carlsson, A. *J. Neural Transm.* **1993**, *94*, 11-19.
- [38] Perachon, S.; Schwartz, J.-C.; Sokoloff, P. *Eur. J. Pharmacol.* **1999**, *366*, 293-300.
- [39] Lorenc-Koci, E.; Wolfarth, S. *Eur. J. Pharmacol.* **1999**, *385*, 39-46.
- [40] Galatsis, P. *Annu. Rep. Med. Chem.* **1996**, *31*, 317.
- [41] Galatsis, P. *Annu. Rep. Med. Chem.* **1997**, *32*, 341.
- [42] Maj, J.; Rogoz, Z.; Skuza, G.; Kolodziejczyk, K. *Eur. J. Pharmacol.* **1997**, *324*, 31-37.
- [43] Iida, M.; Miyazaki, I.; Tanaka, K.; Kabuto, H.; Iwata-Ichikawa, E.; Ogawa, N. *Brain Res.* **1999**, *838*, 51-59.
- [44] Coldwell, M.C.; Boyfield, I.; Brown, T.; Hagan, J.J.; Middlemiss, D.N. *Br. J. Pharmacol.* **1999**, *127*, 1697-1702.
- [45] van Vliet, L.A.; Rodenhuis, N.; Wikstrom, H.; Pugsley, T.A.; Serpa, K.A.; Meltzer, L.T.; Heffner, T.G.; Wise, L.D.; Lajiness, M.E.; Huff, R.M.; Svensson, K.; Haenen, R.M.M.; Bast, A. *J. Med. Chem.* **2000**, *43*, 3549-3557.
- [46] Avenell, K.Y.; Boyfield, I.; Hadley, M.S.; Johnson, C.N.; Nash, D.J.; Riley, G.J.; Stemp, G. *Bioorg. Med. Chem. Lett.* **1999**, *9*, 2715-2720.
- [47] Avenell, K.Y.; Boyfield, I.; Coldwell, M.C.; Hadley, M.S.; Healy, M.A.M.; Jeffrey, P.M.; Johnson, C.N.; Nash, D.J.; Riley, G.J.; Scott, E.E.; Smith, S.S.; Stacey, R.; Stemp, G. Thewlis, K.M. *Bioorg. Med. Chem. Lett.* **1998**, *8*, 2859-2864.
- [48] Austin, N.E.; Avenell, K.Y.; Boyfield, I.; Branch, C.L.; Coldwell, M.C.; Hadley, M.S.; Jeffrey, P.; Johns, A.; Johnson, C.N.; Nash, D.J.; Riley, G.J.; Smith, S.A.; Stacey, R.C.; Stemp, G.; Thewlis, K.M.; Vong, A.K.K. *Bioorg. Med. Chem. Lett.* **1999**, *9*, 179-184.
- [49] Reavill, C.; Taylor, S.G.; Wood, M.D.; Ashmeade, T.; Austin, N.E.; Avenell, K.Y.; Boyfield, I.; Branch, C.L.; Cilia, J.; Coldwell, M.C.; Hadley, M.S.; Hunter, A.J.; Jeffrey, P.; Jewitt, F.; Johnson, C.N.; Jones, D.N.C.; Medhurst, A.D.; Middlemiss, D.N.; Nash, D.J.; Riley, G.J.; Routledge, C.; Stemp, G.; Thewlis, K.M.; Trail, B.; Vong, A.K.K.; Hagan, J.J. *J. Pharmacol. Exp. Ther.* **2000**, *294*, 1154-1165.
- [50] Stemp, G.; Ashmeade, T.; Branch, C.L.; Hadley, M.S.; Hunter, A.J.; Johnson, C.N.; Nash, D.J.; Thewlis, K.M.; Vong, A.K.K.; Austin, N.E.; Jeffrey, P.; Avenell, K.Y.; Boyfield, I.; Hagan, J.J.; Middlemiss, D.N.; Reavill, C.; Riley, G.J.; Routledge, C.; Wood, M. *J. Med. Chem.* **2000**, *43*, 1878-1885.
- [51] Sautel, F.; Griffon, N.; Sokoloff, P.; Schwartz, J.-C.; Launay, C.; Simon, P.; Costentin, J.; Schoenfelder, A.; Garrido, F.; Mann, A.; Wermuth, G. *J. Pharmacol. Exp. Ther.* **1995**, *275*, 1239-1246.
- [52] Schoemaker, H.; Claustre, Y.; Fage, D.; Rouquier, L.; Chergui, K.; Curet, O.; Oblin, A.; Gonon, F.; Carter, C.; Benavides, J.; Scatton, B. *J. Pharmacol. Exp. Ther.* **1997**, *280*, 83-97.
- [53] Bolton, D.; Boyfield, I.; Coldwell, M.C.; Hadley, M.S.; Healy, M.A.; Johnson, C.N.; Markwell, R.E.; Nash, D.J.; Riley, G.J.; Scott, E.E.; Smith, S.A.; Stemp, G.; Wadsworth, H. *Bioorgan. Med. Chem. Lett.* **1996**, *6*, 1233-1236.
- [54] Bolton, D.; Boyfield, I.; Coldwell, M.C.; Hadley, M.S.; Johns, A.; Johnson, C.N.; Markwell, R.E.; Nash, D.J.; Riley, G.J.; Scott, E.E.; Smith, S.A.; Stemp, G.; Wadsworth, H.J.; Watts, E.A. *Bioorg. Med. Chem. Lett.* **1997**, *7*, 485-488.
- [55] Patel, S.; Patel, S.; Marwood, R.; Chapman, K.L.; Emms, K.L.; Freedman, S.B. *Br. J. Pharmacol.* **1995**, *114*, 347P.
- [56] Whetzel, S.Z.; Shih, Y. H.; Georgic, L.M.; Akunne, H.C.; Pugsley, T.A. *J. Neurochem.* **1997**, *69*, 2363-2368.
- [57] Millan, M.J.; Audinot, V.; Rivet, J.-M.; Gobert, A.; Vian, J.; Prost, J.F.; Spedding, M.; Peglion, J.L. *Eur. J. Pharmacol.* **1994**, *260*, R3-R5.
- [58] Witkin, J.; Gasior, M.; Acri, J.; Beekman, M.; Thurkauf, A.; Yuan, J.; DeBoer, P.; Wikstrom, H.; Dijkstra, D.; *Eur. J. Pharmacol.* **1998**, *347*, R1-R3.
- [59] Bristow, L.J.; Cook, G.P.; Patel, S.; Curtis, N.; Mawer, I.; Kulagowski, J.J. *Neuropharmacology* **1998**, *37*, 793-802.
- [60] van Vliet, L.A.; Rodenhuis, N.; Dijkstra, D.; Wikstrom, H.; Pugsley, T.A.; Serpa, K.A.; Meltzer, L.T.; Heffner, T.G.; Wise, L.D.; Lajiness, M.E.; Huff, R.M.; Svensson, K.; Sundell, S.; Lundmark, M. *J. Med. Chem.* **2000**, *43*, 2871-2882.
- [61] Millan, M.J.; Peglion, J.L.; Vian, J.; Rivet, J.-M.; Brocco, M.; Gobert, A.; Newman-Tancredi, A.; Dacquet, C.; Bervoets, K.; Girardon, S. *J. Pharmacol. Exp. Ther.* **1995**, *275*, 895-898.
- [62] Cussac, D.; Newman-Tancredi, A.; Pasteau, V.; Millan, M.J. *Mol. Pharmacol.* **1999**, *56*, 1025-1030.
- [63] Millan, M.J.; Gobert, A.; Newman-Tancredi, A.; LeJeune, F.; Cussac, D.; Rivet, J.-M.; Audinot, V.; Dubuffet, T.; Lavielle, G. *J. Pharmacol. Exp. Ther.* **2000**, *293*, 1048-1062.

- [64] Murray, P.J.; Harrison, L.A.; Johnson, M.R.; Robertson, G.M.; Scopes, D.I.C.; Bull, G.R.; Graham, E.A.; Hayes, A.G.; Kilpatrick, G.J.; Den Daas, I.; Large, C.; Sheehan, M.J.; Stubbs, C.M.; Turpin, M.P. *Bioorg. Med. Chem. Lett.* **1995**, *5*, 219-222.
- [65] Yuan, J.; Chen, X.; Brodbeck, R.; Primus, R.; Braun, J.; Wasley, J.W.F.; Thurkauf, A. *Bioorg. Med. Chem. Lett.* **1998**, *8*, 2715-2718.
- [66] Wustrow, D.; Belliotti, T.; Glase, S.; Kesten, S.R.; Johnson, D.; Colbry, N.; Rubin, R.; Blackburn, A.; Akunne, H.; Corbin, A.; Davis, M.D.; Georgic, L.; Whetzel, S.; Zoski, K.; Heffner, T.; Pugsley, T.; Wise, L. *J. Med. Chem.* **1998**, *41*, 760-771.
- [67] Akunne, H.C.; Zoski, K.T.; Davis, M.D.; Cooke, L.W.; Meltzer, L.T.; Whetzel, S.Z.; Shih, Y.H.; Wurstrow, D.J.; Wise, L.D.; MacKenzie, R.G.; Georgic, L.M.; Heffner, T.G.; Pugsley, T.A. *Neuropharmacology* **2000**, *39*, 1211-1221.
- [68] Corbin, A.E.; Meltzer, L.T.; Ninteman, F.W.; Wiley, J.N.; Christoffersen, C.L.; Wustrow, D.J.; Wise, L.D.; Pugsley, T.A.; Heffner, T.G. *Neuropharmacology* **2000**, *39*, 1197-1210.
- [69] Pilla, M.; Perachon, S.; Sautel, F.; Garrido, F.; Mann, A.; Wermuth, C.G.; Schwartz, J.-C.; Everitt, B.J.; Sokoloff, P. *Nature* **1999**, *400*, 371-375.
- [70] Scheideler, M.A.; Martin, J.; Hohlweg, R.; Rasmussen, J.; Naerum, L.; Ludvigsen, T.S.; Larsen, P.J.; Korsgaard, N.; Crider, A.M.; Ghosh, D.; Cruse, S.F.; Fink-Jensen, A. *Eur. J. Pharmacol.* **1998**, *342*, 153-161.
- [71] Song, X.; Crider, A.M.; Cruse, S.F.; Ghosh, D.; Klein-Stevens, C.; Liang, L.; Scheideler, M.A.; Varming, A.; Sotofte, I. *Eur. J. Med. Chem.* **1999**, *34*, 487-503.